

A NEW 8-HYDROXYFLAVONOL FROM *LARREA TRIDENTATA*

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## INTRODUCTION

In a continuation of our investigation of the flavonoids of *Larrea* species (Zygophyllaceae), we report here the isolation and structure determination of gossypetin 3,7-dimethyl ether (**1**), the third 8-hydroxyflavonol obtained from a hexaploid population of *Larrea tridentata* Cav. from Mojave Co., Arizona, the other two 8-hydroxyflavonols previously isolated from this species being gossypetin 3,7,3'-trimethyl ether and herbacetin 3,7-dimethyl ether [1]. In addition to these 8-hydroxyflavonols, 10 other flavonoids were previously identified from this species: kaempferol, its 3-methyl and 3,7-dimethyl ethers and 3-rhamnoglucoside, isorhamnetin, and quercetin and its 3-methyl and 3,7,3'-trimethyl ethers, 3-glucoside and 3-rhamnoglucoside [2,3].

The new 8-hydroxyflavonol (**1**) was isolated as orange crystals (from MeOH);  $\lambda_{\max}$  MeOH 264sh, 278, 300sh, 344, 378 nm., the band Ia absorption at 378 nm is typical for 8-hydroxyflavonols and a similar absorption was observed for the two other 8-hydroxyflavonols (see above) isolated from the same plant. The new compound exhibited the same bluish color on paper under UV light as these two substances, both of which contain 3,7-methoxyl and 5,8-hydroxyl substitution. The color of **1** changed to yellow-green with ammonia (under UV light) supporting the presence of a free 4'-hydroxyl group. The presence of a B-ring *o*-dihydroxyl group was supported by a hypsochromic shift (22 nm) of band Ia in the  $\text{AlCl}_3$ -HCl spectrum in comparison with the  $\text{AlCl}_3$  spectrum, and by a bathochromic shift (9 nm) of band I in  $\text{NaOAc-H}_3\text{BO}_3$  spectrum in comparison with band I in the MeOH spectrum [4a]. A large bathochromic shift (60 nm) of band

Ia in the  $\text{AlCl}_3$ -HCl spectrum (compared with the MeOH spectrum) appears to be typical for the presence of 5,8-hydroxyl groups in 3-*O*-substituted flavonols [1]; in any case, this result eliminated the possibility of oxygenation at C-6 [4b].

The mass spectrum of **1** exhibited a molecular ion at  $m/e$  346 (51% intensity relative to the base peak) corresponding to  $\text{C}_{17}\text{H}_{14}\text{O}_8$ , that is, for a flavone with four hydroxyl and two methoxyl groups. An M-1 peak at  $m/e$  345 (29%) supported the presence of an 8-hydroxyl group since the loss of a proton from an 8-hydroxyflavonol is a major fragmentation process. An intense peak (60%) at  $m/e$  303 corresponded to a fragment ion ( $\text{M-COMe}$ )<sup>+</sup> typical for either a 3-, 6-, or 8-methoxyl group [5-11]; however, on the basis of the data presented above, **1** must contain a 3- rather than a 6- or 8-methoxyl function [5-13]. A very intense peak at  $m/e$  183 (75%) and a peak at  $m/e$  182 (49%) suggested the presence of a methoxyl group in the A-ring; a base peak at  $m/e$  137 (B-ring plus CO) suggested the presence of a dihydroxyl system in the B-ring (see Fig. 1).

The NMR spectrum of the TMS ether of **1** in  $\text{CCl}_4$  showed a singlet at  $\delta$  3.90 for six protons supporting the presence of two methoxyl groups at the 3- and 7-positions (8-methoxyl groups appear at  $\delta$  3.75-3.8 in many flavonols); a singlet at  $\delta$  6.23 for an H-6 proton; a doublet ( $J=8$  Hz) at  $\delta$  6.84 for the H-5' proton; a double doublet ( $J=8$ , and 2 Hz) at  $\delta$  7.67 for H-6', and a doublet ( $J=2$  Hz) at  $\delta$  7.77 for H=2', supported the presence of a 3',4'-dioxygenated B-ring. The NMR spectrum of the TMS ether of **1** in  $\text{C}_6\text{D}_6$  showed two singlets at  $\delta$  3.27 ( $\Delta + 0.63$  ppm shift; typical for a 7-methoxyl group) and at  $\delta$  3.83

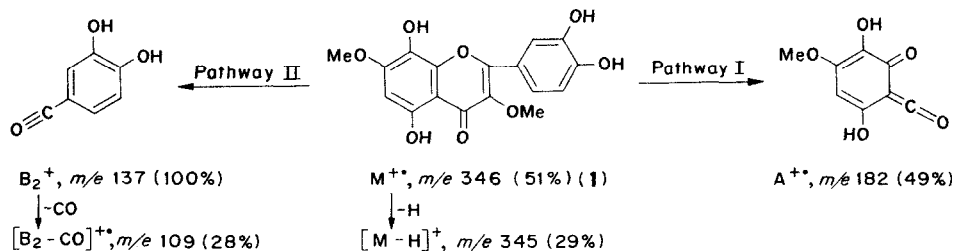


Fig. 1. MS fragmentations of gossypetin 3,7-dimethyl ether.

( $\Delta + 0.07$  ppm; typical for a 3-methoxyl group) [14].

The above spectral findings indicated that compound **1** is a new 8-hydroxyflavonol corresponding to gossypetin 3,7-dimethyl ether.

#### EXPERIMENTAL

NMR spectra were measured at 60 MHz with tetramethylsilane as an internal standard.

Air-dried and ground leaf material from a hexaploid population of *Larrea tridentata* (collected in Mojave Co., Arizona, on July 1971 by Barbara Timmermann) was extracted with 85% aq MeOH; a voucher (B.T., n.s.) is deposited in the University of Texas at Austin Herbarium. The extract was filtered and evaporated down to an aq soln which was extracted with Et<sub>2</sub>O. The ethereal extracts were combined, evaporated and taken to dryness *in vacuo*; 12 g of syrup were chromatographed over polyamide (560 g packed in Egger's solvent, CHCl<sub>3</sub>-MeOH-MeCOEt-2,4-pentanedione, 20:10:5:1. Using Egger's solvent for elution, fractions of about 20 ml each were collected. Rechromatography of fractions 424-454 on a second column of polyamide with MeOH gave orange crystals of gossypetin 3,7-dimethyl ether after recrystallization from MeOH.

Gossypetin 3,7-dimethyl ether (**1**): MS peaks not noted in text: 317 (M-HCO; 9%), 182 (A<sup>+</sup>; 49%), 173 (M<sup>2+</sup>; 20%), 121 (48%), 109 (B<sup>+</sup>; 28%); *R<sub>f</sub>* values: 0.07 (Benzene-HOAc-H<sub>2</sub>O; 6:7:3 upper phase) and 0.38 (25% HOAc); UV in MeOH: 264, 278, 300sh, 344, 378; NaOMe: 264, 396, 457sh (dec.); AlCl<sub>3</sub>: 258sh, 284, 316, 420, 460; AlCl<sub>3</sub>-HCl: 271sh, 285, 312, 361, 438; NaOAc: 263sh, 276, 301sh, 343, 395; NaOAc-H<sub>3</sub>BO<sub>3</sub>: 267, 301sh, 387 (Proc. II). NMR spectral data are described in the text.

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#### REFERENCES

1. Sakakibara, M., Timmermann, B. N., Nakatani, N., Waldrum, H. and Mabry, T. J. (1975) *Phytochemistry* **14**, 849.
2. Chirikdjian, J. J. (1973) *Z. Naturforsch.* **28C**, 32.
3. Chirikdjian, J. J. (1973) *Sci. Pharm.* **41**, 206; (1974) *Pharmazie* **29**, 292.
4. (a) Mabry, T. J., Markham, K. R. and Thomas, M. B. (1970) *The Systematic Identification of Flavonoids*, pp. 13. Springer-Verlag, Heidelberg; (b) Mears, J. A. and Mabry, T. J. (1972) *Phytochemistry* **11**, 411.
5. Kingston, D. G. I. (1971) *Tetrahedron* **27**, 2691.
6. Bowie, J. H. and Cameron, D. W. (1966) *Aust. J. Chem.* **19**, 1627.
7. Joshi, B. S. and Kamat, V. N. (1969) *Ind. J. Chem.* **7**, 636.
8. Bohlmann, F. and Zdero, C. (1967) *Tetrahedron Letters* 3239.
9. Brieskorn, C. H. and Michel, H. (1968) *Tetrahedron Letters* 3447.
10. Nielsen, J. G. (1970) *Tetrahedron Letters* 803.
11. Mabry, T. J. and Markham, K. R. (1975) in *The Flavonoids* (J. B. Harborne, T. J. Mabry and H. Mabry, editors) Chapt. 3. Chapman and Hall, London.
12. Nielsen, J. G. and Moller, J. (1970) *Acta Chem. Scand.* **24**, 2665.
13. Bowie, J. H. and White, P. Y. (1967) *J. Chem. Soc. (C)*, 1933.
14. Rodriguez, E., Carman, N. J. and Mabry, T. J. (1972) *Phytochemistry* **11**, 409.